CYANOBACTERIA MODULATED CHANGES AND ITS IMPACT ON BIOREMEDIATION OF SALINE-ALKALINE SOILS

VENUS SINGH* AND DV SINGH

Department of Botany, Udai Pratap Autonomous College, Varanasi - 221002, Uttar Pradesh, India

Key words: Cyanobacteria, Halotolerance, Bioremediations, Usar soils

Abstract

Saline-alkaline (Usar) soils have high pH and undesirable salts on their surface. A halotolerant, heterocystous and nitrogen fixing cyanobacterium *Nostoc calcicola* Breb. BREB grow successfully on saline-alkaline soils of Eastern Uttar Pradesh. Soil pot experimentation has been conducted in laboratory condition to study the reclamation of saline-alkaline soils collected from investigated site. Both sterilized and natural soils were taken in earthen pots to observe the changes in soil properties inoculated with cyanobacteria and gypsum. In such treated soils significant decrease in pH, ECe and Na⁺ have been observed with cyanobacterial application. There also occurs a significant increase in organic carbon. Experimental reclamation of such soils has been found with cyanobacteria with or without gypsum. *N. calcicola* + gypsum seem to be a suitable combination for reclamation of saline-alkaline soils. This study highlights that a biological amendment with halotolerant cyanobacteria and gypsum in combination shows better option for bioremediation of saline-alkaline (Usar soils).

Introduction

Increasing soil salinity, high soluble salts, sodicity and high exchangeable sodium all are serious land degradation issues worldwide (Vanessa *et al.* 2009). Saline-alkaline soils/salt affected soils (commonly known as Usar soils in India occupy more than 7 million hectare (Mha) land in India (NRSA 2003).

A scientific approach for practicing crop cultivation and to enhance crop productivity in saline/alkaline soil is the reclamation (amelioration) of soil salinity. The reclamation of such soils basically requires the replacement of exchangeable sodium by calcium and thereafter leaching of the exchanged sodium. Conventionally it is done by the approach of addition of gypsum (CaSO₄, 2H₂O) or pyrite (FeSO₄) followed by leaching of excess salts by flooding or extensive irrigation. Biological reclamation through cyanobacteria is another approach. Singh (1961) suggested that cyanobacteria could be used to reclaim alkaline soils because they grow successfully on saline/alkaline soils where most plants fail to grow. Pandey *et al.* (2005), Jaiswal *et al.* (2010) and Murtaza *et al.* (2011) have also suggested the role of cyanobacteria in reclamation of saline-alkaline soils. The objective of the present investigation was to explore the role of halotolerant cyanobacteria in bioremediation of saline-alkaline soils and to study their establishment and influence on soil properties.

Materials and Methods

Soil samples were collected from investigated site (village Amra) is located 20 km away in south-west from Udai Pratap Autonomus College in the district Varanasi ($25^{\circ}20'$ N, $83^{\circ}00'$ E) in the months of April i.e. driest season when very sparse patches of cyanobacterial population were there. The surface soil samples were taken from 0 - 15 cm depth from six different places of the

^{*}Author for correspondence: <singh.venus23@gmail.com>.

area under investigation. Samples were taken in triplicate using soil sampler (auger). Collected soil samples were processed and used for estimation. Before and after inoculation of cyanobacterial species, soil pH and ECe were measured in a soil/water saturation extracts with pH meter and conductivity bridge, respectively following the methods of Richards (1954). Water soluble ions like-Ca²⁺ (Versenate method), Na⁺ (Flame photometrically), organic carbon and sodium adsorption ratio (SAR, mathematically) were also determined as described by Richards (1954). Two experimental sets were prepared, one with natural soil and other with sterilized soil under laboratory condition ($30 \pm 2^{\circ}$ C and light intensity 25 µE/m²/sec).

Allen and Arnon (1955) sterilized medium having pH 9.5 has been used for maintaining the culture. Experimental soils (300 g) were kept in earthen pots (8 cm in height and 20 cm in diameter) and gypsum (powder) was mixed (200 mg/100 g of soil). Natural (unsterilized) as well as sterilized (1.05 kg /cm² pressure for 15 minute) soil sample were used for the experiment. After flooding with sterilized distilled water for 15 days, the cyanobacterial species like *Nostoc calcicola* Breb., *Calothrix* spp., *Anabeana* spp. (inoculum density of 20 µg protein/ml) were inoculated in pots. Four different sets of pot experimentation were done with both the sterilized soils. Pots of Set-1 were taken as control, Set - 2 were added with gypsum only, Set -3 comprised of gypsum + *N. calcicola* and Set - 4 was added with gypsum + mixture of cyanobacteria (*viz; Calothrix* spp., *N. calcicola* BREB, *Anabeana* spp.) and observations were made for 3 months (May - July year needed).

Algal samples were scraped from upper 1 cm surface by using sterilized implements and the cyanobacteria present in the sample were identified with the help of Desikachary (1959) and Rippka *et al.* (1979). Enumeration of cyanobacterial species were done by Pandey *et al.* (1995). The analysis of variance was performed which indicated a significant effect of gypsum on cyanobacterial population. The effect of sterilization was also statistically significant at 5%.

Results and Discussion

Table 1 shows the properties of the soil under observation before inoculation and Table 2 presents the succession of cyanobacterial genera both in natural and sterilized soils. In natural soils (unsterilized) all the cyanobacteria as reported in the Table 2 were observed but, in sterilized condition, only inoculated genera (*Calothrix* spp., *Nostoc calcicola, Anabeana* spp.) were reported.

pН	10.5	—
ECe (dS/m)	6.5	
O.C. (%)	0.10	
Ca^{2+} (Me/l)	3.5	
Na ⁺ (Me/l)	49.0	

Table 1. Characteristics of the saline - alkaline soil before inoculation.

Fig. 1 (a - e) shows the alternation of soil properties by cyanobacterial inoculation in natural soil. All the treatments show decrement of pH, ECe and Na^+ but gypsum in combination with cyanobacteria shows higher decrement than gypsum alone. The same pattern of decrement was also found in case of sterilized soil as in natural soil (Fig. 2).

Application of gypsum and *N. calcicola* have shown lowering of pH and ECe than control pots. A considerable decrease in pH and ECe was also observed in the mixture of gypsum and *N. calcicola*. By comparing the data studied above, application of *N. calcicola* + gypsum seems to be superior over the gypsum + mixture of cyanobacterial form *viz., Calothrix* spp., *N. calcicola* and *Anabaena* spp. for biological reclamation of saline-alkaline soil.

The pattern of change in ECe and organic 'C' after treatment in natural as well as sterilized soils has been shown in Figs 1 b, c and 2 b, c, respectively. In conditions (both natural and sterilized), gypsum and cyanobacteria resulted in enrichment of soil with organic carbon and reduction in ECe.



Fig. 1. Effect of cyanobacteria in altering soil properties in natural soil after 3 months of pot experiment. * Indicates significance from control.

Table 2. Distribution pattern of cyanobacterial species in experimental pots.

Forms		Natural soil	Pot numbers		Sterilized soil			
	1	2	3	4	1	2	3	4
Oscillatoria sp.	Α	А	А	А	-	-	-	-
Microcoleus sp.	Α	Α	А	А	-	-	-	-
Phormidium sp.	Α	-	А	А	-	-	-	-
<i>Lyngbya</i> sp.	Α	А	А	А	-	-	-	-
*Calothrix sp.	А	В	В	С	-	-	-	С
*N. calcicola	Α	В	D	D	-	-	D	D
*Anabaena sp.	А	В	В	В	-	-	-	В
Scytonema sp.	А	А	В	В				

Pot 1 - control, 2 - gypsum, 3 - gypsum + *N.calcicola*, 4 - gypsum + mixture of cyanobacteria (*Calothrix* species, *Nostoc calcicola*, *Anabeana* species).

* Inoculated species A - taxon present, B - common species, C - important, D - dominant.

Soil salinity is a pervasive problem especially in India and the reclamation of these soils is of massive practical importance to reinstate millions of acres of such barren lands once again under the plough (Jaiswal *et al.* 2010). It is clear from the Table 2 that heterocystous cyanobacteria (*Calothrix* sp., *Nostoc calcicola, Anabaena* sp.) were the more frequent inhabitants of the saline-alkaline soils than the non-heterocystous forms and since they have the property to add organic matter and nitrogen to the soil, they could be useful for starting plant succession in barren saline



Fig. 2. Effect of cyanobacteria in altering soil properties in sterilized soil after 3 months of pot experiment. *Indicates significance from control.

soils (Ali and Sandhu 1972). Water leaching a pre-requisite for the reclamation of such soils accompanied by addition of organic matter and chemicals like gypsum or pyrites has also been recommended amendments (Singh 1961, Pandey *et al.* 2005). Due to the high pH of saline-alkaline soils, most likely as a result of Na₂CO₃, the addition of gypsum provides a source of Ca²⁺ which precipitates as CaCO₃ and Ca(HCO₃)₂, resulting in a decrease in soil pH. In addition to decrease in soil pH, the addition of gypsum also leads to proton generation and thus a further reduction in pH occurs (Chorom and Rengasamy 1997). By the addition of gypsum in soil, there occurs an increment of Ca²⁺ and parallel decrement of Na⁺ (depicted in Figs 1 d, e and 2 d, e) which may be due to exchange of Na⁺ by Ca²⁺. In reclamation of saline-alkaline soil, the conversion of Na⁺ clay into Ca²⁺ clay and leaching of excess Na⁺ occurs. Following the addition of gypsum and halotolerant cyanobacterial (*N. calcicola*) to sodic and saline soil, soil microbial biomass (SMB) and respiration rate increased despite adverse soil environmental conditions (Vanessa *et al.* 2009). The addition of gypsum decreases the pH ~ 10.5 - 9.5 in natural soil and 10.5 - 9.7 in sterilized soil. Following the incorporation of gypsum + *Nostoc calcicola* and gypsum

+ mixture cyanobacteria together, the ECe decreases (Figs 1, 2 b). The untreated saline soils (control) were highly alkaline and having high ECe. A significant improvement in soil properties has also been observed after cyanobacterial growth. Alkaline soils with high pH values and Na⁺ content favour the growth of diazotrophic cyanobacteria with a consequent decrease in pH. The decrease in pH is also evident from our findings (Figs 1a and 2a) which are in line with the result of Singh (1961) who reported fall in soil pH from 9.2 to 7.5. Certain organic metabolites produced by cyanobacterial activities are also released in the soils which are responsible for maintaining the fertility of soil year after year (Ladha and Reddy 1995). The presence and succession of diazotrophic cyanobacteria in alkaline soil was reported by Singh (1961) and Singh *et al.* (2014). Frequent observation has been made for the organisms regarding cell characteristics, in that filament with, dead cells, spores and cells with granular body have been observed indicating last stage of their growth cycle. Keeping such views in mind the observation has been made for 3 months. Since gypsum itself is a source of calcium and presence of mucilage in such organisms are responsible for increase in calcium accumulation in soil.

Of the three inoculated genera viz, Anabaena spp., N. calcicola and Calothrix spp., N. *calcicola* were found to be the most dominant forms in all the experimental earthen pots followed by Calothrix and Anabaena spp. (Table 2). Nostoc is known to be one of the most versatile diazotrophic cyanobacterial genera, observed in all types of environments, existing both in free living and symbiotic state. The members of this genus are capable of a variety of modes of C and N nutrition, which adapts them to diverse ecological habitats (Prasanna et al. 2008). N. calcicola adapts synthesis of organic osmoregulatory substances/compatible solutes and improvement in nitrogen assimilation and photosynthetic O₂ evolution which leads to its increased tolerance towards salinity stress (Borowitzka 1986). Moreover, compared to the algae, cyanobacteria belonging to the genus Nostoc offer another advantage: the improvement of total-N soil content, owing to the capability to fix atmospheric nitrogen (Obana et al. 2007). Jaiswal et al. (2010) also suggested N. calcicola as bioameliorating agent for saline-alkaline soil. Addition of Nostoc calcicola biomass to the saline/alkaline soils decreased the pH content and hence improved soil properties. The dominance of *N. calcicola* in saline/alkaline soils may be due to its salt tolerance, which suggests that N. calcicola could be a better phycotechnological approach for soil reclamation.

In conclusion, the role of cyanobacteria in bioremediation of saline-alkaline soil cannot be ignored. A combination of halotolerant cyanobacterium (*N. calcicola*) and gypsum is a considerable option for saline-alkaline/Usar soil reclamation and it should also be a matter of great concern in future experiments related to soil reclamation.

References

Allen MB, and Arnon DI 1955. Studies on nitrogen fixing blue-green algae. I. Growth and nitrogen fixation by *Anabaena cylindrica* Lemm. Plant Physiol. **30**: 366-372.

Ali S, and Sandhu GR 1972. Blue-green algae of the saline soils of the Punjab. Oikos. 23: 268-272.

- Borowitzka LA 1986. Osmoregulation in blue-green algae. Progress in Phycological Research 4: 245-256.
- Chorom M, and Rengasamy P 1997. Carbonate chemistry, pH and physical properties of an alkaline sodic soil as affected by various amendments. Aus. J. Soil Res. **35**: 149-161.

Desikachary TV 1959. Cyanophyta. Indian Council of Agricultural Research. New Delhi, India.

- Jaiswal P, Kashyap AK, Prasanna R, and Singh PK 2010. Evaluating the potential of *N. calcicola* and its biocarbonate resistant mutant as bioameleorating agents for 'usar' soil. Ind. J. Microbiol. **50**: 12-18.
- Ladha JK, and Reddy PM 1995. Extension of nitrogen fixation to rice: necessity and possibilities. Geol J. **35**: 363-372.
- Murtaza B, Murtaza G, Zia-ar-Rehman M, Ghafoor A, Abubakar S, and Sabir M 2011. Reclamation of saltaffected soils using amendments and growing wheat crop. Soil Environ. **30**(2): 130-136.

NRSA 2003. Mapping of salt affected soils. National Remote Sensing Agency, Hyderabad, India.

- Obana S, Miyamoto K, Morita S, Ohmori M, and Inubushi K 2007. Effect of *Nostoc* sp. on soil characteristics, plant growth and nutrient uptake. J. Appl. Phycol. **19**: 641-646.
- Pandey KD, Kashyap AK, Gupta RK 1995. Nutrient status, algal and cyanobacterial flora of six fresh water streams of Schirmacher Oasis, Antarctica. Hydrobiologia 299:83-91.
- Pandey KD, Shukla PN, Giri DD, and Kashyap AK 2005. Cyanobacteria in alkaline soil and the effect of cyanobacteria inoculation with pyrite amendments on their reclamation. Biol. Fertil. Soils 41: 451-457.
- Prasanna R, Jaiswal P, Kaushik BD 2008. Cyanobacteria as potential options for environmental sustainability-promises and challenges. Ind. J. Microbiol. 48: 89-94.
- Richards LA 1954. Diagnosis and improvement of saline and alkali soils. Agricultural Handbook no. 60. United States Department of Agriculture, Washington, DC.
- Rippka R, Deruelles J, Waterbury JB, Herdman M, Stanier RY 1979. Generic assignments, strain histories and properties of pure cultures of cyanobacteria. J. Gen. Microbiol. 11: 1-61.
- Singh RN 1961. Role of blue green algae in nitrogen economy of Indian agriculture. Indian Council of Agricultural Research New Delhi.
- Singh Veenus, Pandey KD, Singh DV 2014. Cyanobacterial succession in saline-alkaline soils with seasonal variation in soil properties. Advances in Plant Sciences **27**(I): 119-123.
- Vanessa NL, Wong Ram C D, Richard SBG 2009. Carbon dynamics of sodic and saline soils following gypsum and organic material additions: A laboratory incubation. Appl. Soil Ecol. 41: 29-40.

(Manuscript received on 31 May, 2015; revised on 13 September, 2015)